



Research

A SCITECHNOL JOURNAL

A combined use of Chondrocytes Micro Grafts (CMG) Mixed with Platelet Rich Plasma (PRP) in Patients Affected by Pinch Nose Deformity

Gentile P^{1*}, Scioli MG², Bielli A², Orlandi A³ and Cervelli V⁴

Abstract

Background: The combined use of autologous chondrocytes micro-grafts (CMG) mixed with Platelet Rich Plasma (PRP) is an alternative that opens a new era in this field.

Materials and Methods: At the Department of Plastic and Reconstructive Surgery, University of Rome "Tor Vergata", Italy, 15 patients, affected by pinch nose deformity underwent nasal alar reconstruction with chondrocytes micro-grafts gently poured onto PRP in solid form. A CT scan control was performed after 12 months. Pearson's Chi-square test was used to investigate difference in cartilage density (CD) between native and newly formed cartilage.

Results: The constructs of CMG-PRP subcutaneously injected resulted in cartilage tissue with adequate central nutritional perfusion. Postoperative follow-up evaluation has shown optimal aesthetic results and improvement of nasal obstruction. These composite grafts provide functional support to the alar cartilages, usually collapsed in pinch nose deformity because of excessive resection during previous surgery.

Conclusion: This report demonstrated that chondrocytes micro-grafts derived from nasal septum poured onto PRP in solid form are a useful method for cartilage regeneration in patients affected by external nasal valve collapse.

Keywords

Chondrocytes auto-graft; Cartilage regeneration; Cartilage tissue engineering

Introduction

For therapeutic cartilage regeneration, the use of chondrocytes micrografts (CMG) mixed with platelet-rich plasma (PRP) has not been studied for recapitulating chondrogenesis. PRP has been demonstrated to be effective in the treatment of soft tissue defects and pattern hair loss [1-4]. It has been demonstrated that three-dimensional culture system in type I collagen scaffold and the addition of multiple growth factors contained in PRP, in the culture

medium induced proliferation and a robust chondrogenesis of adult stem cells *in vitro* [5,6]. Direct mixing of chondrocytes with PRP leads to shrinking and deformed cartilage formation *in vivo* [7] owing to poor mechanical stability and rapid degradability. In particular the use of chondrocytes micrografts represents a micro-invasive procedure and grafts are more flexible to fill the lesions with various shapes. Today the main problem in transferring the experimental protocols of tissue engineering in the routine clinical practice is the identification of accessible sites where an adequate amount of stem cells are collected [8,9]. In addition, the need to specifically define technical procedure and its safety is an essential factor, as occurring for stem cell application in breast reconstruction and soft tissue defects [10-13].

Although within the human body there are several "niches" inhabited by a significant number of stem cells [14-16], often these are not easy to access. The nasal septum represents a niche housing chondrocytes easily accessible without limited morbidity of the anatomical site after collection of the micrografts. Ruikai Ba et al. [17] developed the cell bricks technique, which cultured a chondrocyte sheet and cut such a cell-Extra Cellular Matrix complex into multiple small fragments (cell bricks). They found that chondrocyte bricks significantly inhibited vascular infiltration into PRP gels and slowed their degradation, thus maintaining the framework and shape of the PRP grafts [18]. They hypothesized that the cell brick-enriched PRP gel could be an ideal injectable niche for adults stem cells, which is expected to regenerate biological cartilage tissues with persistent cartilaginous phenotype, less deformation and uniform histological structure [18].

Currently, in this study, we investigated the *in vivo* performance of chondrocytes in cell brick-enriched PRP gels, and evaluated the persistence of a stable chondrogenic phenotype.

Chondrocytes can be obtained by enzymatic digestion and/or mechanical disaggregation: in the first method, the cartilage tissue is harvested under sterile conditions, digested with appropriate enzymes, and then the resulting cell suspensions are seeded in culture dishes containing a special medium supplemented with necessary additives and then incubated. Finally, the resulting colonies are sub-cultured before confluence and the cells are stimulated to differentiate. In the second procedure, chondrocytes are isolated by the mechanical centrifugation of cartilage (Rigenera Method) in which the cartilage is treated as any other connective tissue subjected to grafts, with a phase of collection and a phase of mechanical disaggregation of the tissue without manipulating the matrix. Rigenera protocol produces millions of viable micro-grafts and filters them with a cut-off of 50 microns, in order to promote the discharging of old differentiated cells and the enrichment of young progenitors cells contained within the cartilage.

Materials and Methods

Patients

A total of 15 patients aged 23-67 years, affected by external nasal valve collapse, the so-called pinched nose deformity, were treated from January 2014 to September 2015 at the Plastic and Reconstructive Surgery Department of "Tor Vergata" University, Rome.

*Corresponding author: Pietro Gentile, MD, PhD, Via Courmayeur, No. 102, 00135 Rome, Italy, Tel: +39-3388515479; E-mail: pirogentile2004@libero.it

Received: July 06, 2016 Accepted: July 27, 2016 Published: August 01, 2016

The preoperative study was carried out through a complete clinical examination of the nasal pyramid and of the nasal cavities through anterior rhinoscopy, a photographic examination in three projections and CT scans.

The authors cut the septum in strips during rhynoplasty. The strips were gently collected and dissociated using Rigenera System (HBW srl, Turin, Italy) in 1.2 ml of physiologic solution. After a specific centrifugation the suspension was harvested from the system and infiltrated onto PRP gel.

The product obtained (PRP gel mixed with chondrocytes) was applied on the defect in external nasal valve collapse (Figure 1F). After 12 months follow-up radiographic examinations were done with the use of CT scans.

PRP preparation

PRP was prepared with approval of the transfusional service from a small volume of blood (18 ml) according to the method of the Cascade-Selphyl-Esforax system (Figure 2A-C) (Aesthetic Factors, LLC, Wayne, NJ, www.selphyl.com), with modifications, and from 60 ml of blood according to the P.R.L. Platelet Rich Lipotransfert system (CORIOS Soc. Coop, San Giuliano Milanese, Italy, www.corios.it), with modifications, using PRP alone (C-punt; Biomed Device, Modena, Italy, www.biomeddevice.it) without the combined use of stromal vascular fraction cells. In the Cascade-Selphyl-Esforax procedure, we used centrifuge 1,100 g for 10 minutes (Figure 2A); in the P.R.L. Platelet Rich Lipotransfert system and C-Punt System, we used centrifuge 1,200 RPM for 10 minutes). Then, autologous PRP, not activated (9 ml) obtained by Cascade-Selphyl procedure, was switched with 10-ml tubes containing Ca²⁺. The authors obtained PRP in solid form (Figure 1A) after the second spin of PRP activated with Ca²⁺.

Autologous PRP, not activated, obtained by the C-Punt Biomed Device procedure after centrifugation (20 ml), was inserted in a light selector device. At the end of the procedure, 9 ml of PRP was harvested.

Micro-grafts chondrocytes preparation

Micro-grafts of chondrocytes were prepared from a Rigeneracons® (CE certified Class I). After the extraction of the nasal septum (Figure 3A and B) during rhynoplasty, the authors cut the septum (Figure 3C and D) in strips (2 mm × 2 mm) (Figure 3E and F). The strips was dissociated using Rigeneracons® System (HBW srl, Turin, Italy) (Figure 2D and E) in 1.2 ml of physiologic solution (Figure 2F). After one minute of centrifugation at 80RPM the cellular suspension was harvested from the system and infiltrated onto PRP gel (Figure 1A).

The aim is to disaggregate a small piece of cartilage (septum cartilage strips) and select a cell population with a size of 50 micron. These cell populations were analyzed by the authors in hystological evaluation section.

Cytospin and alcian-PAS staining

Chondrocytes, isolated by Rigenera System (HBW srl, Turin, Italy), were made to adhere on a glass slide by cytospin, then Alcian-PAS staining was performed (Ventana-Roche Diagnostics, Milan, Italy). Positive cells were counted in the total area under a light microscope (Eclipse E600, Nikon, Japan) and microphotographs captured by DXM1200F Digital camera (Nikon) using ACT-1 software (Nikon).

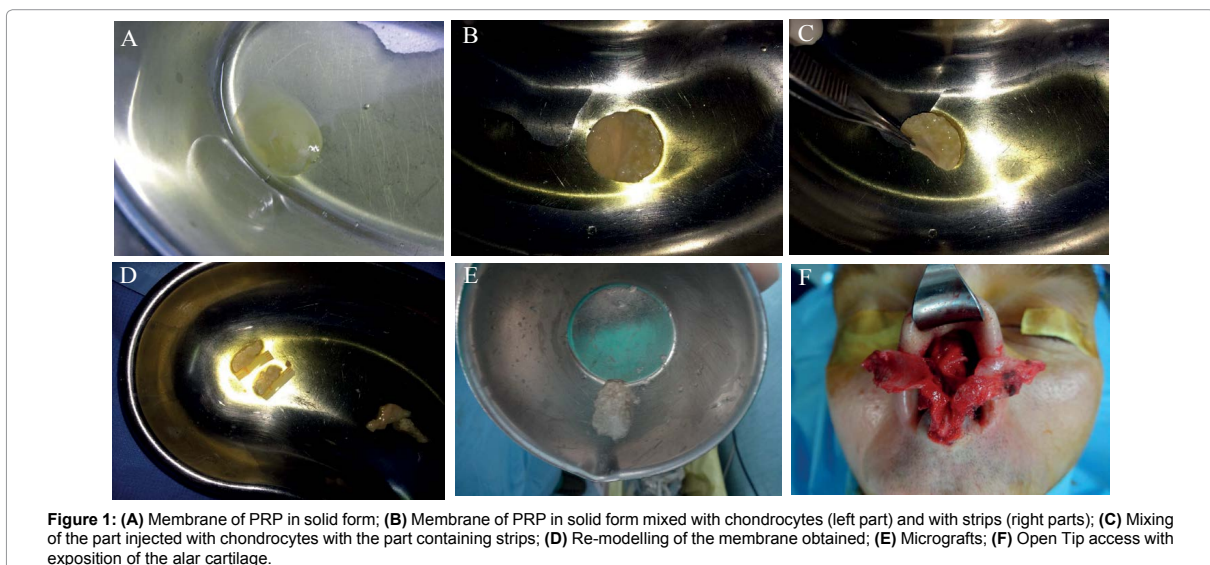
Histological evaluation

Excisional tissue was obtained from four randomly selected patients after 6 months from surgery. Microscopic evaluation of routine Haematoxylin-Eosin stained paraffin embedded sections [19] was performed to morphologically analyze cartilage tissue.

Results

Clinical and Instrumental observation

In this study, we reported the results with the use of composite autologous chondrocytes micro-grafts with PRP, as exemplified by the following case. Nasal obstruction was very marked in these patient; external nose analysis showed external nasal valve collapse



(Figure 4A), deficit of supratip projection. Anterior rhinoscopy showed a deviated nasal septum and bilateral stenosis of the internal valves. The composite graft, was applied on the external nasal valve collapse, in the alar cartilage side by fixing with absorbable stitches. Postoperative follow-up evaluation has shown optimal aesthetic results (Figure 4B) and improvement of nasal obstruction. These composite grafts provide functional support to the alar cartilages, usually collapsed because of excessive resection during previous surgery. Transcolumellar open-tip access was necessary to allow for better visualization of the valve collapse, alar cartilage and for fixation of the cartilaginous structures, to allow for placement of unexposed absorbable stitches.

In the CT scans it shows the pre-operative situation in comparison with the regenerated site in post-operative image. Figure 1B-E shows the preparation.

Histological observation

Chondrocytes isolated by Rigenera System, were stained with Alcian-PAS and counted under a light microscope. The mean of cell yield was $9,088.30 \pm 788.86$ cells/ml of digestion.

In addition, microscopic analysis of excisional fragments, in two patients, showed the persistence of healthy cartilage tissue surrounded by a fibrous connective tissue layer in which newly formed capillaries spread and penetrate into cartilage.

Discussion

The regenerative surgery opens new challenges for correction of nasal deformity. An injectable approach for cartilage regeneration could meet today's demand for micro-invasive surgery [20]. Zhu et al. [18] showed that cell bricks (CB) formed by fragmented chondrocyte macro-aggregates could stabilize the PRP gel efficiently *in vivo* and support chondrogenesis with stable morphology. They found that such an injectable complex could support chondrogenesis of bone marrow mesenchymal stem cells (BMSCs) *in vivo*, and demonstrated that such a completely biological graft could meet the requirement of nasal augmentation. The most important finding in this study

is that BMSCs embedded in chondrocyte brick-enriched PRP gel underwent persistent chondrogenesis, and hypertrophic translation was prevented [18]. In addition, BMSCs are capable to promote angiogenesis through the secretion of growth factors, in particular VEGF [21]. Angiogenesis is a crucial event for tissue regeneration [22], as well as for tumor growth [23,24].

This finding provides a micro-invasive approach for cartilage regeneration, indicating the promising future of cell bricks for clinical application. A surprising phenomenon presented in study of Ba, et al. [17] is that BMSCs mixed into CB-PRP gel guided fast angiogenesis in BMSC regions and prevented central necrosis of the whole graft. Their previous experiment encountered the problem that chondrocytes mixed into CB-PRP gels resulted in obvious necrosis in the interior of constructs, which could be attributed to the robust anti-angiogenic potential of chondrocytes. Their results confirmed that BMSCs in CB-PRP gels presented higher VEGF expression than chondrocytes in CB-PRP gels, which was regarded to contribute the angiogenesis in BMSCs regions in PRP grafts.

Here we demonstrated that Rigenera System is a useful method to isolate human chondrocytes and when cells were injected with PRP *in vivo* in patients affected by nasal valve collapse.

The constructs of chondrocytes micro-grafts-PRP that were subcutaneously injected resulted in a persistent cartilage tissue with appropriate morphology, adequate central nutritional perfusion without central necrosis or ossification, and further augmented nasal dorsum without obvious contraction and deformation.

In addition, microscopic analysis of excisional fragments showed the persistence of healthy cartilage tissue with the formation of new capillaries penetrating into cartilage.

Appropriate scaffolds combined with growth factors can significantly improve the survival and differentiation of the transplanted stem/progenitor cells [6,19,25-31].

In a study of Brunelli et al. [25], the micro-grafts obtained by dental pulp poured onto collagen sponge are useful method for bone regeneration in atrophic maxilla. Dental stem/progenitor cells



Figure 2: (A) Cascade Centrifuge; (B) Tube extracted by Cascade kit for membrane preparation containing CaCl; (C) Phase of preparation; (D) Rigenera Centrifuge; (E) Rigenera Kit; (F) Syringe with 1ml of solution containing chondrocytes.

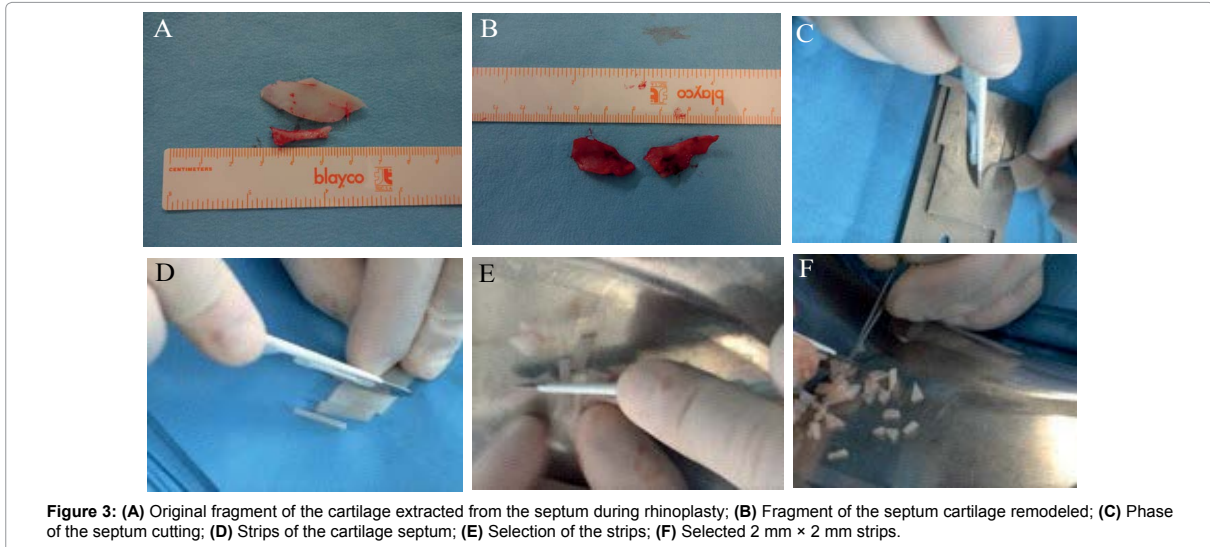


Figure 3: (A) Original fragment of the cartilage extracted from the septum during rhinoplasty; (B) Fragment of the septum cartilage remodeled; (C) Phase of the septum cutting; (D) Strips of the cartilage septum; (E) Selection of the strips; (F) Selected 2 mm × 2 mm strips.

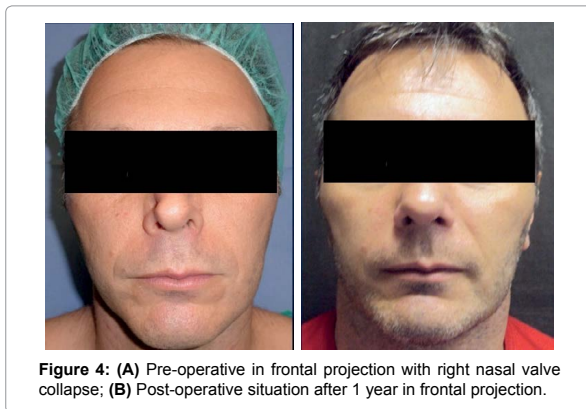


Figure 4: (A) Pre-operative in frontal projection with right nasal valve collapse; (B) Post-operative situation after 1 year in frontal projection.

(DPSCs) collected from dental pulp can be differentiated in vitro and then transplanted with biomaterial scaffolds into the host without immunologic rejection [32,33]. Graziano et al. [34] observed DPSCs performances on different scaffolds, such as PLGA or poly (lactic-co-glycolic acid) 85:15, hydroxyapatite chips (HA), and titanium. Results showed that stem cells exerted a different response, depending on the different type of textured surface [35]. Thus, cells cultured on the concave textured surface had better cell-scaffold interactions and were induced to secrete factors that, due to their autocrine effects, quickly lead to osteogenic differentiation, bone tissue formation, and vascularization.

References

- Cervelli V, Gentile P, Scioli MG, Grimaldi M, Casciani CU, et al. (2009) Application of platelet-rich plasma in plastic surgery: clinical and in vitro evaluation. *Tissue Eng Part C Methods* 15: 625-634.
- Cervelli V, Garcovich S, Bielli A, Cervelli G, Curcio BC, et al. (2014) The effect of autologous activated platelet rich plasma (AA-PRP) injection on pattern hair loss: clinical and histomorphometric evaluation. *Biomed Res Int* 2014: 760709.
- Gentile P, Garcovich S, Bielli A, Scioli MG, Orlandi A, et al. (2015) The Effect of Platelet-Rich Plasma in Hair Regrowth: A Randomized Placebo-Controlled Trial. *Stem Cells Transl Med* 4: 1317-1323.
- Yamada Y, Nakamura S, Ito K, Kohgo T, Hibi H, et al. (2008) Injectable tissue-engineered bone using autogenous bone marrow-derived stromal cells for maxillary sinus augmentation: clinical application report from a 2-6-year follow-up. *Tissue Eng Part A* 14: 2099-1707.
- Cervelli V, Scioli MG, Gentile P, Doldo E, Bonanno E, et al. (2012) Platelet-rich plasma greatly potentiates insulin-induced adipogenic differentiation of human adipose-derived stem cells through a serine/threonine kinase Akt-dependent mechanism and promotes clinical fat graft maintenance. *Stem Cells Transl Med* 1: 206-220.
- Scioli MG, Bielli A, Gentile P, Cervelli V, Orlandi A (2015) Combined treatment with platelet-rich plasma and insulin favors chondrogenic and osteogenic differentiation of human adipose-derived stem cells in 3D collagen scaffold. *J Tissue Eng Regen Med*.
- Wu W, Chen F, Liu Y, Ma Q, Mao T (2007) Autologous injectable tissue-engineered cartilage by using platelet-rich plasma: experimental study in a rabbit model. *J Oral Maxillofac Surg* 65: 1951-1957.
- Sinanani AC, Hunt NP, Lewis MP (2004) Human adult craniofacial muscle-derived cells: Neural-cell adhesion-molecule (NCAM; CD56)-expressing cells appear to contain multipotential stem cells. *Biotechnol Appl Biochem* 40: 25-34.
- Papaccio G, Graziano A, d'Aquino R, Graziano MF, Pirozzi G, et al. (2006) Long-term cryopreservation of dental pulp stem cells (SBP-DPSCs) and their differentiated osteoblasts: A cell source for tissue repair. *J Cell Physiol* 208: 319-325.
- Gentile P, Orlandi A, Scioli MG, Di Pasquali C, Bocchini I, et al. (2012) A comparative translational study: the combined use of enhanced stromal vascular fraction and platelet-rich plasma improves fat grafting maintenance in breast reconstruction. *Stem Cells Transl Med* 1: 341-351.
- Gentile P, Orlandi A, Scioli MG, Di Pasquali C, Bocchini I, et al. (2012) Concise review: adipose-derived stromal vascular fraction cells and platelet-rich plasma: basic and clinical implications for tissue engineering therapies in regenerative surgery. *Stem Cells Transl Med* 1: 230-236.
- Bielli A, Scioli MG, Gentile P, Agostinelli S, Tarquini C, et al. (2014) Adult adipose-derived stem cells and breast cancer: a controversial relationship. *Springerplus* 3: 345.
- Bielli A, Scioli MG, Gentile P, Cervelli V, Orlandi A (2015) Adipose Tissue-Derived Stem Cell Therapy for Post-Surgical Breast Reconstruction—More Light than Shadows. *Adv Clin Exp Med* 24: 545-548.
- Bianco P, Robey PG (2001) Stem cells in tissue engineering. *Nature* 414: 118-121.

15. Calvi LM, Adams GB, Weibrecht KW, Weber JM, Olson DP, et al. (2003) Osteoblastic cells regulate the haematopoietic stem cell niche. *Nature* 425: 841-846.
16. Zhang J, Niu C, Ye L, Huang H, He X, et al. (2003) Identification of the haematopoietic stem cell niche and control of the niche size. *Nature* 425: 836-841.
17. Ba R, Wei J, Li M, Cheng X, Zhao Y, et al. (2015) Cell-bricks based injectable niche guided persistent ectopic chondrogenesis of bone marrow-derived mesenchymal stem cells and enabled nasal augmentation. *Stem Cell Res Ther* 6: 16.
18. Zhu J, Cai B, Ma Q, Chen F, Wu W (2013) Cell bricks-enriched platelet-rich plasma gel for injectable cartilage engineering – an in vivo experiment in nude mice. *J Tissue Eng Regen Med* 7: 819-830.
19. Pisano C, De Cesare M, Beretta GL, Zuco V, Pratesi G, et al. (2008) Preclinical profile of antitumor activity of a novel hydrophilic camptothecin, ST1968. *Mol Cancer Ther* 7: 2051-2059.
20. Hausamen JE (2001) The scientific development of maxillofacial surgery in the 20th century and an outlook into the future. *J Maxillofac Surg* 29: 2-21.
21. Kinnaid T, Stabile E, Burnett MS, Lee CW, Barr S, et al. (2004) Marrow-derived stromal cells express genes encoding a broad spectrum of arteriogenic cytokines and promote in vitro and in vivo arteriogenesis through paracrine mechanisms. *Circ Res* 94: 678-685.
22. Sun X, Altalhi W, Nunes SS (2015) Vascularization strategies of engineered tissues and their application in cardiac regeneration. *Adv Drug Deliv Rev* 96: 183-194.
23. Cassinelli G, Zuco V, Petrangolini G, De Cesare M, Tortoreto M, et al. (2012) The curative efficacy of namitecan (ST1968) in preclinical models of pediatric sarcoma is associated with antiangiogenic effects. *Biochem Pharmacol* 84: 163-171.
24. Ciuffreda L, Di Sanza C, Cesta Incani U, Eramo A, Desideri M, et al. (2012) The mitogen-activated protein kinase (MAPK) cascade controls phosphatase and tensin homolog (PTEN) expression through multiple mechanisms. *J Mol Med (Berl)* 90: 667-679.
25. Brunelli G, Motroni A, Graziano A, D'Aquino R, Zollino I (2013) Sinus lift tissue engineering using autologous pulp micro-grafts: A case report of bone density evaluation. *J Indian Soc Periodontol* 17: 644-647.
26. Graziano A, d'Aquino R, Laino G, Papaccio G (2008) Dental pulp stem cells: A promising tool for bone regeneration. *Stem Cell Rev* 4: 21-26.
27. Srisawasdi S, Pavasant P (2007) Different roles of dexamethasone on transforming growth factor-beta1-induced fibronectin and nerve growth factor expression in dental pulp cells. *J Endod* 33: 1057-1060.
28. Iohara K, Zheng L, Ito M, Tomokiyo A, Matsushita K, et al. (2006) Side population cells isolated from porcine dental pulp tissue with self-renewal and multipotency for dentinogenesis, chondrogenesis, adipogenesis, and neurogenesis. *Stem Cells* 24: 2493-2503.
29. Ueno A, Yamashita K, Miyoshi K, Horiguchi T, Ruspita I, et al. (2006) Soluble matrix from osteoblastic cells induces mineralization by dental pulp cells. *J Med Invest* 53: 297-302.
30. Hosoya A, Nakamura H, Ninomiya T, Hoshi K, Yoshida K, et al. (2007) Hard tissue formation in subcutaneously transplanted rat dental pulp. *J Dent Res* 86: 469-474.
31. Otaki S, Ueshima S, Shiraishi K, Sugiyama K, Hamada S, et al. (2007) Mesenchymal progenitor cells in adult human dental pulp and their ability to form bone when transplanted into immunocompromised mice. *Cell Biol Int* 31: 1191-1197.
32. d'Aquino R, Graziano A, Sampaoli M, Laino G, Pirozzi G, et al. (2007) Human postnatal dental pulp cells co-differentiate into osteoblasts and endotheliocytes: A pivotal synergy leading to adult bone tissue formation. *Cell Death Differ* 14: 1162-1171.
33. Graziano A, d'Aquino R, Laino G, Proto A, Giuliano MT, et al. (2008) Human CD34+stem cells produce bone nodules in vivo. *Cell Prolif* 41: 1-11.
34. Graziano A, d'Aquino R, Cusella-De Angelis MG, De Francesco F, Giordano A, et al. (2008) Scaffold's surface geometry significantly affects human stem cell bone tissue engineering. *J Cell Physiol* 214: 166-172.
35. d'Aquino R, De Rosa A, Laino G, Caruso F, Guida L, et al. (2009) Human dental pulp stem cells: From biology to clinical applications. *J Exp Zool B Mol Dev Evol* 312 B: 408-415.

Author Affiliations

Top


¹Plastic and Reconstructive Surgery Department, University of Rome "Tor Vergata", Rome, Italy

²Institute of Anatomic Pathology, University of Rome Tor Vergata, Italy

³Chief of Anatomic Pathology University of Rome Tor Vergata, Italy

⁴Chief of Plastic and Reconstructive Surgery Department, University of Rome "Tor Vergata", Italy

Submit your next manuscript and get advantages of SciTechnol submissions

- ❖ 50 Journals
- ❖ 21 Day rapid review process
- ❖ 1000 Editorial team
- ❖ 2 Million readers
- ❖ More than 5000 
- ❖ Publication immediately after acceptance
- ❖ Quality and quick editorial, review processing

Submit your next manuscript at ● www.scitechnol.com/submission